



DEVELOPMENT, STANDARDIZATION AND VALIDATION OF HPLC METHOD FOR ESTIMATION OF β -SITOSTEROL IN *VITEX NEGUNDO* SEED EXTRACT

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ABSTRACT

A simple, precise and accurate high-performance liquid chromatographic method has been developed for determination of β -sitosterol in the hydroalcoholic extract of *Vitex negundo* Linn. A Reverse phase HPLC column CrestPack RP C18 (Make: Jasco; 250 mm x 4.6 mm I.D; particle size 5 μ m) was used as stationary phase with a mobile phase comprising of acetonitrile : methanol (70:30) v/v at a flow rate of 1.0 mL min⁻¹ and UV detection at 210nm with a run time of 6 mins. The proposed method was validated for linearity, accuracy, precision, LOD and LOQ. LOD for beta-sitosterol was found to be 0.5 μ g/mL whereas LOQ for the same was found to be 1.5 μ g/mL. The method was found to be linear over the range 1 μ g/mL to 5 μ g/mL with coefficient of regression 0.9732 for betasitosterol. Intra-day and inter-day precision studies showed % RSD to be less than 2 and hence the method was precise. Stock solution stability studies were carried out for beta-sitosterol. Stability samples in methanol after a week revealed that all beta-sitosterol in solution form were stable. The validated HPLC method can be used for a routine quality control analysis and quantitation of β -sitosterol from *Vitex negundo* Linn.

Key words: β -sitosterol, HPLC, nirgundi, validation, *Vitex negundo*, seed extract.

INTRODUCTION

Vitex negundo belonging to family Verbenaceae is a large aromatic shrub distributed throughout the greater part of India up to an altitude of 1500 meter in the Himalayas. The shrub is very common in many part of the country and often occurs gregariously. It is abundant along the banks, rivers in moist situations, and open waste lands. It is widely planted as hedge plant along the roads and between fields. The shrub can be reproduces readily from cutting and as it produces roots, suckers it is useful for planting against soil erosion [1]. The traditional systems of Siddha and Ayurvedic medicine use this plant alone or in combination with other medicinal plants for the treatment of various diseases. It contains various chemical classes such as steroids, flavonoids, glycosides, alkaloids, proteins, tannins and phenolic compounds. One of the important constituent being betasitosterol.

β -Sitosterol is white, waxy phytosterol (plant sterols) with chemical structures similar to that of cholesterol. It is widely distributed in the plant kingdom [2] responsible for reduction of cholesterol level in plasma [3] and improves liver function activity (GDP, GOP) [4].

HPTLC [5-9] and HPLC [2,10,11] methods have also been reported for analysis of β - sitosterol either individually or in combination with other marker compounds. Over the past decade HPLC has been successfully used in the analysis of pharmaceuticals, plant constituents, and biomacromolecules [12]. A major advantage of HPLC is that it has the ability to easily separate a wide variety of chemical mixtures. Hence the present study was undertaken to develop and validate simple, accurate, precise and robust analytical procedure for estimation of β -sitosterol in *Vitex negundo* seed extract by HPLC.

MATERIAL AND METHODS

Collection

Vitex negundo Linn. (Chaste tree) seeds were collected from the local market Borivali, Mumbai, Maharashtra in the month of June 2013. Authentication of the collected material was carried out at Blatter Herbarium St. Xavier's College, Mumbai by Dr. Rajendra D. Shinde, and its identity was confirmed to be *Vitex negundo* Linn. (Chaste tree), family Verbenaceae, with herbarium

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Processing of *Vitex negundo* Seeds

Vitex negundo seeds were garbled and unwanted plant parts, was remove. Seeds were then allowed to dry at room temperature. Dried seeds were ground to powder in mixer and sieved through 20# to get seed powder.

Extraction

Extraction of seeds of *Vitex negundo* was carried in soxhlet extractor using ethanol and water (70:30) as solvent. 100gm of dried and powdered seeds were weighed and packed in filter paper, then extracted with solvent (500ml) at a temperature not exceeding 80°C in a soxhlet apparatus for 72 hrs. After completion of extraction the solvent was evaporated in evaporating dish on the water bath to obtain dry extract. The weight of dry extract was taken to find out extractive value. The extractive value of the hydroalcoholic extract of *V. negundo* was found to be 6.4% w/w.

Quantification of β -sitosterol in *V.negundo* seed extract Instrumentation

The analysis of the drug was carried out on a Jasco LC system equipped with 2089 pump and photodiode array detector (PDA) was used and a Reverse phase HPLC column CrestPack RP C18 (Make: Jasco; 250 mm x 4.6 mm I.D; particle size 5 μ m) was used. The output of signal was monitored and integrated using Chromnav software.

Chemicals and solvents

Milli-Q Water, Acetonitrile (ACN) (HPLC Grade), Methanol (MeOH) (HPLC Grade) was obtained from Merck, Mumbai.

Mobile phase preparation

Prepare a filtered and degassed mixture of Acetonitrile and Methanol in the ratio of 70:30 v/v respectively.

Standard preparation

Accurately weigh and transfer about 10 mg of beta sitosterol into a 10 ml volumetric flask, add 4 ml of Methanol, sonicate to dissolve and dilute to volume with Methanol to yield solutions of 1000 μ g/mL. This solution was further diluted equimolarly to give stock solution of 100 μ g/mL concentration. Further dilutions were made as required with methanol.

Sample preparation

Accurately weigh and transfer about 100 mg of *Vitex negundo* seed extract into a 10 ml volumetric flask, add 4 ml of Methanol, sonicate to dissolve and dilute to volume with Methanol to yield solutions of 10000 μ g/mL.

Chromatographic conditions

CrestPack RP C18 (Make: Jasco; 250 mm x 4.6 mm I.D; particle size 5 μ m) Column was used for analysis at ambient column temperature. The mobile phase was

pumped through the column at a flow rate of 1.0 ml/min. The sample injection volume was 20 μ l. The photodiode array detector was set to a wavelength of 210 nm for the detection and Chromatographic run time was 6 minutes.

RESULTS AND DISCUSSION

1) Method development

Choice of column

Choice of the column was made depending on the physicochemical properties of the drug. Since beta-sitosterol is hydrophobic in nature therefore reverse – phase chromatography was considered to be the best choice. C18 column being more hydrophobic as compared to C8, retention of drug on C18 column was better. Hence C18 column was selected.

Development and optimization of mobile phase

Many mobile phases were initially used however the peaks obtained were broad and retention time was a problem. This was solved by using a different mobile phase ACN: Methanol (70:30) This mobile phase composition resulted in symmetric peak and good retention time.

Detection system

The presence of chromophore makes UV-visible detection more suitable for determination of β -sitosterol using PDA Detector. Selection of λ_{max} was done based on spectrophotometric scan of compounds. Betasitosterol showed appropriate absorption at 210 nm. PDA detector enabled to determine peak identity and purity.

Method validation

The developed RP-HPLC method extensively validated for the determination betasitosterol Content using the following Parameters.

a) Linearity and Range

The calibration curve was prepared by injecting various concentrations of the standard solutions. The calibration plot was found to be linear in the range 1 μ g/mL to 5 μ g/mL with a correlation coefficient 0.9732 for betasitosterol.

b) Limit of Detection

The limit of detection (LOD) was obtained by successively decreasing the concentration of beta-sitosterol as long as a signal-to-noise ratio of 3:1 appeared. The LOD for beta-sitosterol was found to be 0.5 μ g on column (volume of injection was 20 μ l).

c) Limit of Quantification

The limit of quantification (LOQ) for beta-sitosterol was found to be 1.5 μ g on column (volume of injection was 20 μ l).

d) Precision and Accuracy

Precision was reported in terms of coefficient of variance (%CV) over the range of quantitation for a single experiment in which standards were assayed in replicate

(Intraday) and for a series of experiments in which standards were assayed in over several experiments (Interday).

e) Specificity and Selectivity

The method was quite selective for beta-sitosterol since there was no other interfering peak around their retention time. The baseline did not show any significant peak.

f) Stability

The stability studies was evaluated for three different concentration i.e. 2 µg/mL, 3 µg/mL and 4 µg/mL. For stock solution stability, solutions corresponding to the above three concentrations were stored at refrigerated temperature (2-8°C) for a week. Stability was calculated by comparing the results of six

replicate injections of stored solutions with fresh samples.

Application of Validated Method to The Analysis Of Samples

Sample Preparation

10 mg of dried extract was weighed and diluted suitably with methanol for HPLC analysis.

Content in plants

The extracts of samples were injected in the concentration that would fall within the linearity of standards injected. For calculation, from the calibration curve we get concentration for extract. Amount of betasitosterol present in the hydro-ethanolic extract of *V.negundo* seeds is approximately 0.7266 % w/w. Therefore amount of betasitosterol present in the seeds is 0.0465% w/w.

Table 1. HPLC Method Parameters

HPLC System	Jasco HPLC system
Pump	Jasco PU-2089 Plus Quaternary Gradient HPLC Pump
Detector	Jasco MD-2018 Plus PDA Multiwavelength Detector
Integrator	Chromnav software
Stationary phase	Jasco CrestPak C18 (4.6×250mm,5µm)
Mobile phase	ACN :Methanol (70:30% v/v)
Detection wavelength	210 nm
Flow rate	1 mL/min
Sample size	20 µl

Table 2. Linearity For β-sitosterol standard

Conc. (µg/mL)	Lin 1	Lin 2	Lin 3	Lin 4	Lin 5	MEAN	± S.D.	%C.V
1.0	231962	231957	231985	231876	231979	231951.8	43.92835072	0.018938569
2.0	235790	235697	235756	235617	235845	235741	87.68409206	0.037195096
3.0	237600	237587	237698	237513	237660	237611.6	71.18497035	0.029958542
4.0	239418	239517	239470	239498	239118	239404.2	164.2808571	0.068620708
5.0	241514	241570	241598	241878	242174	241746.8	277.1771996	0.114655995

Table 3. Intraday Precision And Accuracy For β-sitosterol

Sr. No	Conc. Per spot		
	2	3	4
1	2.1	3.2	4.12
2	2.05	3.04	3.96
3	2.16	2.93	3.94
4	2.12	2.87	4.08
5	1.97	3.16	4.02
6	1.89	3.14	4.1
Mean	2.048333333	3.056666667	4.036666667
SD	0.101472492	0.133666251	0.075277265
%CV	4.953905232	4.372941692	1.86483729
%Accuracy	102.4166667	101.8888889	100.9166667

Table 4. Interday Precision And Accuracy For β-sitosterol

CONC	2	3	4
1	2.1	3.2	4.12
2	2.05	3.04	3.96
3	2.16	2.93	3.94
4	2.12	2.87	4.08
5	1.97	3.16	4.02

6	1.89	3.14	4.1
7	2.16	3.15	4.11
8	1.98	3	3.93
9	2.17	2.97	3.94
10	2.02	2.89	4.15
11	1.99	3.06	4.01
12	1.86	3.1	4.08
Mean	2.039166667	3.0425	4.036666667
SD	0.105352942	0.111202109	0.079696394
%CV	5.166470397	3.654958396	1.97431198
% Accuracy	101.9583333	101.4166667	100.9166667

Table 5. Stability For β -sitosterol

Sr. No.	Conc. of Fresh Solution			Conc. of Stability Solution		
	2 $\mu\text{g/mL}$	3 $\mu\text{g/mL}$	4 $\mu\text{g/mL}$	2 $\mu\text{g/mL}$	3 $\mu\text{g/mL}$	4 $\mu\text{g/mL}$
1	2.1	3.2	4.02	2.06	3.05	4.01
2	2.05	3.04	3.96	1.98	3	3.93
3	2.06	2.97	3.94	2.07	2.97	3.97
4	2.11	2.97	4.08	2.02	2.89	4.1
5	1.97	3.06	4.02	1.99	3.06	4.01
6	1.93	3.09	4.1	1.87	3.02	4.08
MEAN	2.037	3.055	4.02	1.9983	2.9983	4.017
\pm S.D	0.072	0.085	0.063	0.072	0.062	0.064
% CV	3.536	2.814	1.573	3.628	2.082	1.603
% STABILITY	101.833	101.833	100.5	99.917	99.944	100.416

Table 6. HPLC Calibration Data

Conc.	Area
1 $\mu\text{g/ml}$	231962
2 $\mu\text{g/ml}$	235790
3 $\mu\text{g/ml}$	237600
4 $\mu\text{g/ml}$	239418
5 $\mu\text{g/ml}$	241514
extract (10000 $\mu\text{g/ml}$)	395612

Fig 1. Structure of β -sitosterol

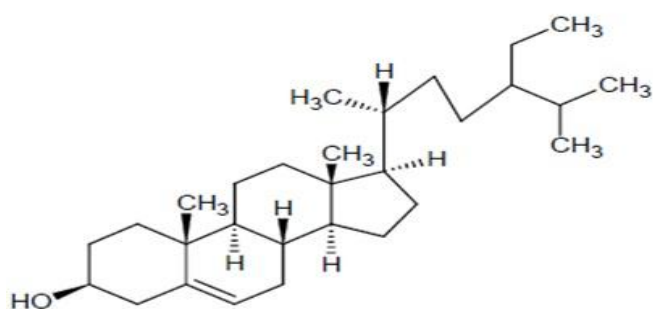


Fig 2. A typical HPLC Chromatogram showing the Peak of β -sitosterol standard

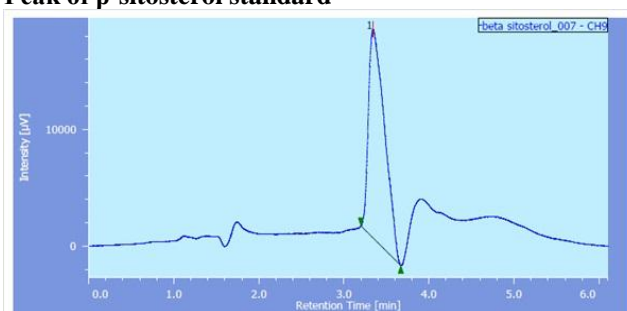


Fig 3. Linearity For β -sitosterol standard

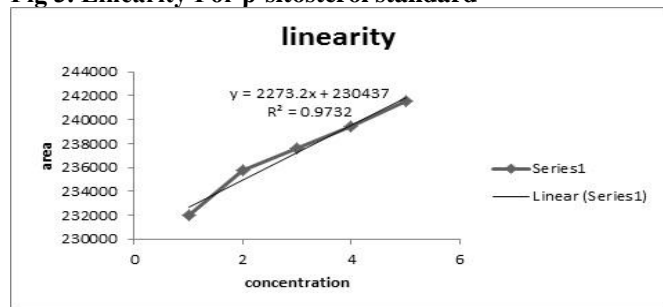
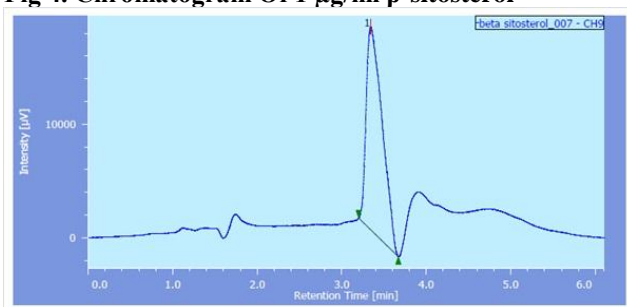
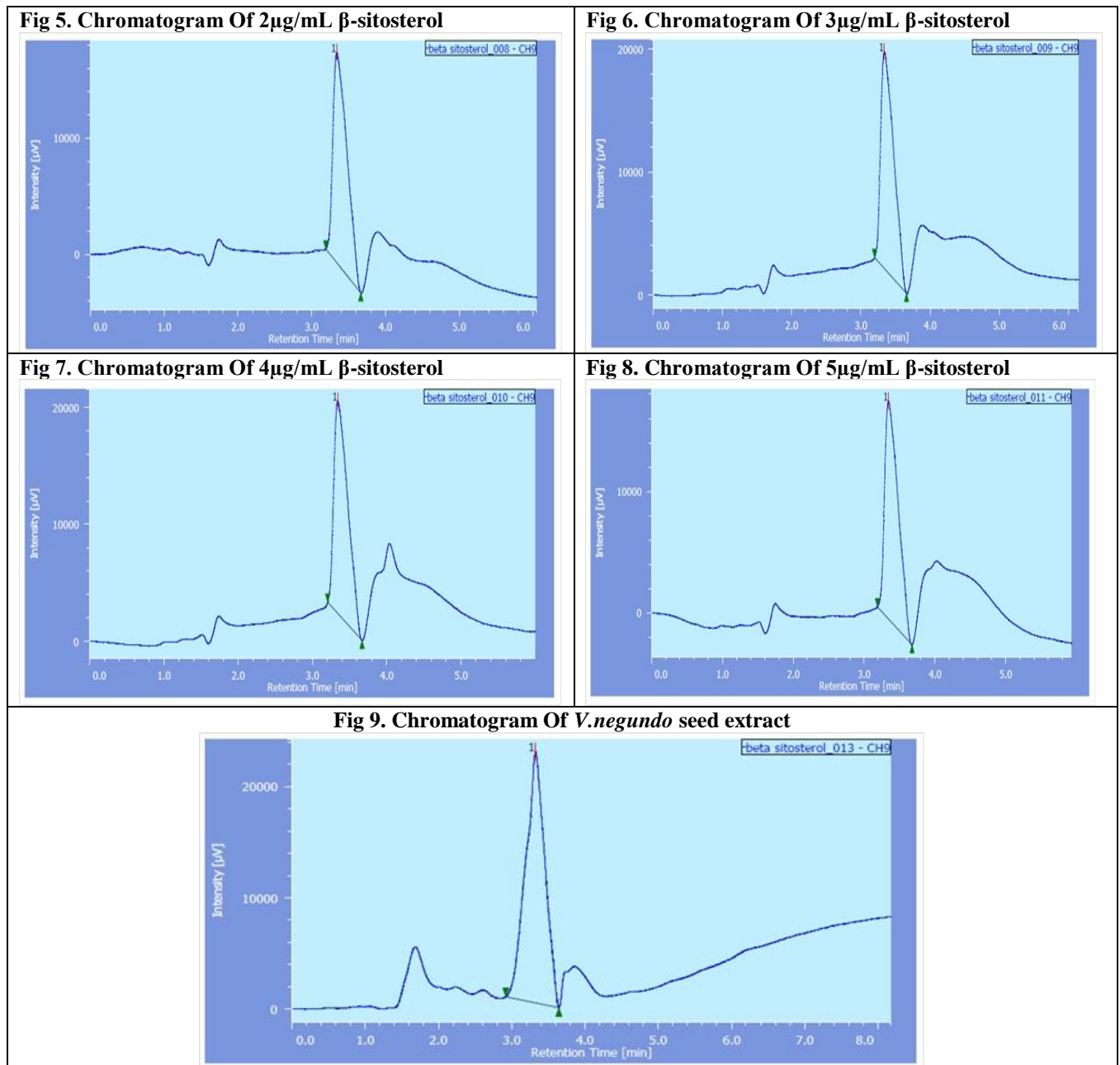


Fig 4. Chromatogram Of 1 $\mu\text{g/ml}$ β -sitosterol





CONCLUSION

We have developed a fast, simple and reliable analytical method for determination of betasitosterol in *Vitex negundo* seed extract using HPLC. It is very fast, with good reproducibility and good response. Validation

of this method was accomplished, getting results meeting all requirements. The method is simple, reproducible, with a good accuracy and precision. It allows reliable analysis of betasitosterol in *Vitex negundo* seed extract.

REFERENCES

1. Gupta M, Mazumdar UK and Bhawal SR. CNS activity of *Vitex negundo* Linn. in mice. *Indian J Exp Biol*, 37, 1999, 143-146.
2. Kakade AN, Magdum CS. HPLC analysis of β -sitosterol in herbal medicine and vegetable oils. *Int J Pharm & Life Sci (IJPLS)* 3(5), 2012, 1666-1669.
3. MacLatchy DL, Van Der Kraak GJ. The phytoestrogen betasitosterol alters the reproductive endocrine status of goldfish. *Toxicol Appl Pharmacol*, 134, 1995, 305-312.
4. Zak A, Vecka M, Tvrzicka E, Hruby M, Novak F, Papezova H, Lubanda H, Vesela L, Stankova B. Composition of plasma fatty acids and non-cholesterol sterols in anorexia nervosa. *Physiol Res*, 54, 2005, 443-451.
5. Jirge SS, Tatke PA, Gabhe SY. Development and validation of a novel HPTLC method for simultaneous estimation of Betasitosterol D glucoside and Withaferin A. *Int J Pharm Pharm Sci*, 3(2), 2011, 227-230.

6. Dighe V, Adhyapak S, Tiwari K. Quantitation of β -sitosterol from *Celastrus paniculatus* Willd. using validated high performance thin layer chromatography method. *Int Pharma and Bio Sci*, 2(2), 2011, 117-123.
7. Shailajan S, Shah S, Sayed N. HPTLC method development and validation of a secondary metabolite – β - sitosterol from *Caesalpinia Bonduc* (linn.) Roxb. Emend. Dandy & exell. Seeds. *Int Pharma and Bio Sci*, 1(3), 2010, 1-10.
8. Bumrela SB, Naik SR. Identification of β -carotene and β - sitosterol in methanolic extract of *Dipteracanthus patulus* (Jacq) nees and their role in antimicrobial and antioxidant activity. *Int J Phytomed*, 3, 2011, 204-215.
9. Misar A, Mujumdar AM, Ruikar A, Deshpande NR. Quantification of β -sitosterol from barks of three *Acacia* species by HPTLC. *J Pharm Res*, 3(11), 2010, 2595-2596.
10. Lopez Ortiz CM, Prats Moya MS, Berenguer NV. A rapid chromatographic method for simultaneous determination of β -sitosterol and tocopherol homologues in vegetable oils. *J Food Compos Anal*, 19, 2006, 141-149.
11. Nair VDP, Kanfer I, Hoogmartens J. Determination of stigmasterol, beta-sitosterol and stigmastanol in oral dosage forms using high performance liquid chromatography with evaporative light scattering detection. *J Pharmaceut Biomed*, 41, 2006, 731-737.
12. Kadam AB, Havele SS, Dhaneshwar SR. Validated HPTLC method for simultaneous estimation of lafutidine and domperidone in bulk drug and formulation. *Int J Pharm Pharm Sci*, 4(4), 2012, 221-225.
13. Willey S, Kekare MB and Vaidya V. Development and Validation of High Performance Liquid Chromatographic Method For the Simultaneous Determination of β -Sitosterol and Lupeol in *Vernonia cinerea* Linn. *International Journal of Pharma and Bio Sciences*, 1(3), 2010, 1-5.